Safety and Antiviral Activity of the HCV Entry Inhibitor ITX5061 in Treatment-Naïve HCV-Infected Adults: A Randomized, Double-Blind, Phase 1b Study

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Abstract

Background: HCV entry involves scavenger receptor B1 (SRB1). In vitro, SRB1 inhibition by ITX5061 impedes HCV replication.

Methods: Multicenter study to assess safety/activity of ITX5061 in previously untreated, non-cirrhotic, HCV genotype 1 infected adults. Design included sequential cohorts of 10 subjects with ITX5061 (n=8) or placebo (n=2) to escalate duration (3 to 14 to 28 days) or de-escalate dose (150 to 75 to 25 mg) based on pre-defined criteria for safety and activity (≥4 of 8 subjects with HCV RNA decline ≥ 1 log₁₀ IU/mL).

Results: Thirty subjects enrolled in three cohorts: ITX5061 150 mg/day by mouth for 3 (A150), 14 (B150), and 28 (C150) days. Six subjects had grade ≥3 adverse events (one in placebo); none were treatment related. One of the seven C150 subjects (14.3%, 95% CI 0.7%-55.4%) had ≥ 1 log₁₀ IU/mL decline in HCV RNA (1.49 log₁₀ IU/mL) whereas none of the six placebo, eight A150 or eight B150 subjects showed such decline.

Conclusions: Oral ITX5061 150 mg/day for up to 28 days was safe and well tolerated. In the 28-day cohort, 1 of 7 subjects showed antiviral activity; however, predefined criteria for antiviral activity were not met at the doses and durations studied.

Introduction

Chronic HCV infection is a major cause of liver-related morbidity and mortality due to cirrhosis and hepatocellular carcinoma (HCC).(1) Current HCV therapy is suboptimal due to toxicity, lack of broad efficacy, and poor compliance.(2-4) Although over two dozen drugs are being developed against the replicative enzymes of HCV (e.g., NS3/4A protease and NS5B polymerase), the area of entry inhibitors is largely unexplored.(5) The use of the in vitro full length JFH-1 (HCV genotype 2a) virus replication system as well as the pseudotyped viruses has allowed characterization of HCV entry.(6) While many details remain unclear, cellular receptors necessary for HCV entry include scavenger receptor class B, type1 (SR-B1), cluster of differentiation 81 (CD81), claudin-1, and occludin.(7-10) ITX5061 is a high potency small molecule inhibitor of SR-B1 which is an integral trans-membrane protein found in numerous cell types, including the liver. It is a functionally active cell surface high-density lipoprotein (HDL) receptor and is an important component of the reverse cholesterol transport pathway, mediating selective HDL delivery from plasma HDL to the liver.(11-13) Since inhibition of SR-B1 is associated with increases in plasma HDL, ITX5061 was evaluated for treatment of hypoalphalipoproteinemia. (14;15) While subsequent trials did not support this, ITX5061 was administered to ~ 280 humans with good safety and reproducible pharmacokinetics (unpublished data).

In vitro, ITX5061 differentially binds to SR-B1 expressing cells and specifically inhibits the binding of soluble HCV E2. Further, ITX5061 inhibits HCV cell culture derived (HCVcc) infection at subnanomolar potency, with a therapeutic window > 10,000 fold.(14) In vitro, it acts additively with interferon, RBV, and the protease inhibitor, telaprevir and, based on its unique mechanism of action, no cross-resistance is expected between ITX5061 and HCV polymerase or protease inhibitors.(16)

The objective of A5277 was to assess the safety and antiviral activity of ITX5061 in patients infected with HCV genotype 1 who had not previously received HCV treatment.

Methods

Study design. A5277 was a randomized, double-blind, placebo-controlled study to find a safe dose of ITX5061 with antiviral activity. The trial consisted of 3 parts designed to evaluate anti-HCV activity of ITX5061 over three drug exposure durations: 3 days (Part A), 14 days (Part B), 28 days (Part C). The planned treatment durations were based on mathematical modeling of the effect of blocking HCV entry on viral decay in the blood through the loss of infected hepatocytes, which suggested that 4 weeks might be required to observe a significant decline in viremia (Dr. Alan Perelson, personal communication). Based on this modeling, tempered with the concern for selecting resistant variants during monotherapy, a range of durations were explored from 3 days to 4 weeks in a sequential fashion. Within each part (A, B, and C), up to 3 doses of ITX5061 given by mouth once daily could be tested (150, 75, and 25 mg). Based on the extensive, human safety data, dosing in each part began with the highest ITX5061 dose (150 mg). The rationale for this approach was that if antiviral activity was demonstrated for the high dose at a given duration, then dose de-escalation within the same duration cohort could be performed to establish a minimum effective dose.

The initial cohort, A150, assessed ITX5061 150 mg/day for 3 days in 10 subjects (8 active and 2 placebo). The study design planned for subsequent cohorts to be enrolled based on the safety and antiviral activity observed in previous cohorts. An acceptable safety profile in 8 subjects treated with active drug in each cohort was defined as ≤ 2 subjects with Grade 3 events and none with Grade 4 or Grade 5 (death) events attributed to the study drug; attribution took into account site opinion, other potential causes and timing of the event. Antiviral activity in 8 subjects treated with active drug in each cohort was defined as ≥ 4 subjects with serum HCV RNA ≥ 1 log₁₀ IU/mL decrease from baseline (day 0) at the end of treatment.

After the completion of each dose/duration cohort, the safety and antiviral activity data were reviewed by the Cohort Review Group (protocol leadership team, an independent clinician and the study statisticians) to determine the next steps. With the exception of the statisticians, CRG members were blinded to the treatment assignment. If the safety criteria were not met, additional cohorts would not be

enrolled. If the safety criteria were met, the next cohort to be enrolled would depend on the observed antiviral activity. For the first cohort (150 mg for 3 days, A150), if both safety and antiviral activity criteria were met, the next cohort to enroll would be the next lower dose (75mg) of ITX5061 administered for the same duration; however, if safety was observed but antiviral activity was not demonstrated, the next cohort to enroll would be the same dose of ITX5061 for longer treatment duration (14 days). The decision algorithm for additional cohorts was similar with the exception of cohort C150 (150 mg for 28 days duration). If antiviral activity was not observed in the C150 cohort, no additional cohorts were to be enrolled (**Figure 1**).

Population. The study enrolled men and women between the ages of 18 and 65 years with previously untreated, chronic HCV genotype 1 infection and no significant hepatic fibrosis. Liver fibrosis ≤ METAVIR stage 2 was confirmed by biopsy (within 2 years) or HCV FibroSURE™ (within 1 year).(17) Individuals with decompensated liver disease, hepatocellular carcinoma, Gilbert's and coinfection with HIV-1 and/or hepatitis B virus were excluded. Subjects were also required to have laboratory parameters within the following ranges: Absolute neutrophil count (ANC) ≥ 1000/mm³; Hemoglobin ≥ 12 g/dL for men and ≥ 11 g/dL for women; Platelet count ≥ 120,000/mm³; Alanine aminotransferase (ALT) ≤ 5 x upper limit of normal (ULN); International normalized ratio (INR) < 1.5; Total bilirubin ≤ ULN; Calculated creatinine clearance (CrCl) ≥ 80 mL/min, by the Cockcroft-Gault equation. Subjects who participated in an earlier cohort were not eligible for subsequent cohorts.

Study drug. Both ITX5061 and placebo were formulated as a 25 mL oral solution and stored under refrigerated conditions. ITX5061 was prepared as 150 mg [6 mg/mL], 75 mg [3 mg/mL], and 25 mg [1 mg/mL]) in a vehicle consisting of 20% (w/w) hydroxypropyl-beta-cyclodextrin in 10 mM aqueous citric acid.

HCV virology. Serum HCV RNA levels were assessed prior to, during and following treatment with ITX5061 or placebo. For the 3 day cohorts, HCV RNA was scheduled on treatment days 0, 1, 2 and on post-treatment days 3, 9 and 16. For the 14 day cohorts: HCV RNA was scheduled on treatment days 0, 1, 2, 3, 7, 10, 13 and on post-treatment days 14, 20 and 27. For the 28 day cohorts, HCV RNA was

scheduled on treatment days 0, 1, 2, 3, 7, 10, 14, 21, 27 and on post-treatment days 28, 34 and 41.

Serum HCV RNA quantification was performed at the ACTG Virology Specialty Laboratory using the COBAS® TaqMan® HCV Test, v1.0 (Roche Molecular Systems, Pleasanton, California). For each cohort, HCV RNA measurement was performed simultaneously in batch on all specimens.

Pharmacology. ITX5061 was taken with food. In Part A, pre-dose samples were collected at Day 0 and Day 1 and an intensive pharmacokinetic study was completed on Day 2. In Part B, pre-dose samples were collected on Days 0, 1, 2, 3, 7 and 10 and an intensive pharmacokinetic study was completed on Day 13. In Part C, pre-dose samples were collected on Days 0, 7, 14 and 21 and an intensive pharmacokinetic study was completed on Day 27. The intensive pharmacokinetic study included sampling over an 8-hour period (0 [pre-dose], 15 minutes, 30 minutes, 1, 2, 3, 4, 6, 8 hours) and a 24hour sample collected when the subject returned on the next day. Samples were obtained by venipuncture in EDTA anticoagulant. The whole blood samples were centrifuged at 800 x g. The resulting plasma was transferred into cryovials and stored at -70°C. Shipment of samples was completed overnight on dry ice and was stored at -70°C. Plasma ITX5061 concentrations were determined with a previously reported liquid chromatography/mass spectrometry assay.(18) Mass spectrometric conditions were determined after optimization of both ITX5061 and the deuterated internal standard in a 1:1 mixture of acetonitrile in 0.1% formic acid solution. Precursor ions of ITX5061 and the internal standard were confirmed from the spectra obtained from infusion of aqueous solutions of the analytes. Acceptance criteria for the lower limit of quantification (LLOQ) required a coefficient of variation (CV%) value of ≤20% and the remaining quality controls required a CV% value of ≤15%. Accuracy (% deviation) for the quality controls was within ± 15%. The data were analyzed by noncompartmental analysis using Phoenix 64, WinNonlin 6.3. Linear trapezoidal method was used for computation of area under the concentration-time curve (AUC).

Statistical Methods. The primary endpoint on antiviral activity was binary: HCV RNA decrease $\geq 1 \log_{10}$ IU/mL from baseline (day 0) at the end of treatment. With the study antiviral activity criteria of ≥ 4 subjects with HCV RNA $\geq 1 \log_{10}$ IU/mL decrease in each cohort of 8 active subjects, the probability of

concluding activity within cohort was 89% assuming 65% true proportion with such HCV RNA reduction. The confidence intervals (CI) around proportions were calculated using the Blyth-Still-Casella method. The continuous measures are summarized using median, Q1 (1st quartile) and Q3 (3rd quartile); mean and standard deviation (SD) are also provided for the pharmacokinetic parameters. Wilcoxon Signed Rank tests were used to assess significant changes from baseline in continuous measures, and the relationships between treatment duration and response in continuous measures were assessed using Jonckheere-Terpstra tests. Spearman's rank correlation was used to examine association between continuous measures. Statistical tests were two-sided, and p-value<0.05 was considered significant in each test without adjustment for multiple tests in this exploratory study. Analyses were conducted using SAS version 9.2.

Results

Study population. The study enrolled 30 subjects (24 active; 6 placebo) sequentially in three cohorts with the administration of ITX5061 150 mg daily for 3 days (A150, December 2010 to March 2011), 14 days (B150, May 2011 to July 2011) and 28 days (C150, October 2011 to Jan 2012) at ten US sites (Table 1). Of the 30 subjects, 16 were female (53%); 15 were non-Hispanic black (50%), 9 were non-Hispanic white (30%) and 6 (20%) were Hispanic regardless of race. The median age was 50 years and the median body weight was 88.2 kilograms. At baseline, the median HCV RNA level was 6.21 log₁₀ IU/mL (Q1=5.72, Q2=6.87). The majority (97%) of subjects completed study treatment and follow-up. One subject on active treatment enrolled in C150 stopped ITX5061 after 2 days due to personal reason (not side effects) and discontinued the protocol at day 6. This subject was excluded from antiviral activity and pharmacokinetic analyses.

Safety. Of the 30 enrolled subjects, 6 (A150, n=3; B150, n=2; C150, n=1) were reported to have grade 3 or higher adverse events at or after study initiation; all events were either not treatment related or existed prior to drug/placebo dosing (**Supplemental Table 1**). One subject i who received placebo experienced alcohol withdrawal with an elevated AST level. Four subjects who received ITX5061 had grade 3 conditions prior to dosing: polysubstance abuse (n=2), depression/bipolar disorder (n=1),

chronic back pain (n=1), elevated serum ALT, and leg pain (n=1). New signs/symptoms and laboratory events were of mild to moderate severity (grade 1 and 2, see **Supplemental Table 2),** and no significant changes in liver enzymes, total bilirubin, ALT and AST levels were observed.

Virologic activity and serum HDL levels. None of the 8 subjects who received 150 mg of ITX5061 for 3 days (A150) and 14 days (B150) had a ≥ 1 \log_{10} IU/mL decrease in HCV RNA level at the end of dosing compared to baseline: one-sided 97.5% confidence interval is (0%,34.9%) within cohort. In the 28 day cohort (C150), one of 7 subjects who completed ITX5061 dosing had a decrease in HCV RNA > 1 \log_{10} IU/mL from baseline to the end of dosing (1.49 \log_{10} IU/mI) (Figure 2). This subject was a 63 year old non-Hispanic, black female with baseline HCV RNA level of 5.58 \log_{10} IU/mL and a serum alanine aminotransferase level that was 2.7 times the upper limit of normal; the subject's HCV RNA and serum HDL changes over time are displayed in **Figure 3**. The estimated proportion of subjects with antiviral activity in C150 was 14.3% (two-sided 95% CI: 0.7% to 55.4%). None of the 6 subjects treated with placebo demonstrated antiviral activity. Similarly, none of the actively treated ITX5061 arms demonstrated statistically significant changes in \log_{10} HCV RNA level at the end of treatment compared to baseline (all p-values > 0.05).

The median change in log₁₀ HCV RNA from baseline to the end of treatment among actively treated subjects was: 3 day cohort, 0.06 log₁₀ IU/mL (Q1=– 0.06, Q3=0.17; P=0.46); 14 day cohort, -0.04 log₁₀ IU/mL (Q1= – 0.11,Q3= 0.08; P=0.64); 28 day cohort, 0.03 log₁₀ IU/mL (Q1= – 0.14, Q3= 0.10; P=0.94) (**Table 2**). There was no suggestion of increased viral reduction with longer duration (p=0.43). As a surrogate for SR-B1 blockade, serum total and HDL levels at baseline and at the end-of-treatment were also assessed (**Supplemental Table 3**). The median change in HDL level was: 3 day cohort, 9 mg/dL (Q1=6, Q3=14, P=0.008); 14 day cohort, 2 mg/dL (Q1=– 4, Q3=9, P=0.74); 28 day cohort, 10 mg/d (Q1=–1, Q3=25, P=0.125). Overall, the change in HDL level was not significantly correlated with the change in HCV RNA level, and no statistically significant dose-response relationship was detected in the end of treatment change from baseline in HCV RNA and any lipid level (total and HDL cholesterol).

Pharmacology. As shown in **Figure 4**, the plasma concentration profiles in A150, B150 and C150 yielded similar profiles with a rapid absorption phase achieving a peak concentration within the first 1-2 hours. The peak was then followed by a bi-exponential pattern with an initial decline over the next 8 hours and a more prolonged terminal phase between 12 to 24 hours. Trough concentrations and pharmacokinetic parameters demonstrated a pattern that reflected the long plasma half-life requiring prolonged dosing to reach a steady-state. Table 3 summarizes the plasma concentrations and pharmacokinetic parameters of interest. There was statistically significant trend of shorter half-life and longer treatment duration (p=0.02). No statistically significant associations were observed between pharmacokinetic parameters (AUC, Cmin and Cmax) and changes from baseline at the end of treatment in (a) HCV RNA and (b) HDL. No trends over time were noted to suggest relationship between HCV RNA change and trough concentration, and between HDL change and trough concentration.

Discussion

Advances in the understanding of the biology of HCV has led to the development of very promising antiviral drugs targeted to non-structural proteins with enzymatic function (e.g., NS3/4A protease) or other non-structural proteins involved in replication (e.g., NS5A). Similar advances have been made with respect to deciphering HCV entry into hepatocytes using multiple cellular receptors including scavenger receptor class B, type1 (SRB1). While antiviral strategies designed to inhibit entry including blocking antibodies and small molecules are under investigation, our study is one of the first clinical trials of a putative HCV entry inhibitor in adults with chronic HCV infection. As such, several of our findings are noteworthy in this study evaluating the potential effect of viral entry inhibition in chronically infected patients.

The effect, if any, that inhibition of viral entry has on levels of viremia is unknown. In contrast, the antiviral activity of drugs that effectively inhibit HCV non-structural proteins can be readily detected by monitoring HCV RNA levels over a few days. We evaluated the SRB1 entry inhibitor for up to 28 days at a dose previously shown to raise HDL cholesterol levels in patients. Nonetheless, only one of 23

subjects dosed with ITX5061 for 3, 14 or 28 days demonstrated a greater than 1 log₁₀ reduction in HCV RNA from baseline, and the median HCV RNA level in all cohorts was similar at pretreatment and the end of dosing. The subject with a significant HCV response in the 28 day cohort had a relatively low HCV RNA at baseline and had an increase in HDL cholesterol that was consistent with SRB1 inhibition. However, this individual was similar to other subjects who did not demonstrate viral suppression with respect to ITX5061 exposure and, overall, there was no significant correlation between changes in HDL and HCV RNA levels across the entire study population. Taken together, ITX5061 alone is unlikely to have a significant effect on viremia in chronically infected patients over the dose (150 mg) and durations (up to 28 days) of exposure studied.

One potential explanation for the lack of observed antiviral effect could be that the magnitude of drug concentration achieved at 150 mg daily was inadequate to inhibit HCV entry. However, the plasma pharmacokinetics of ITX5061 were similar to those observed in prior studies and appeared to achieve concentrations that were in excess of the in vitro inhibitory values that had been used to design the dosing regimens. However, ITX5061 is highly bound to plasma proteins in vitro (\sim 99%); thus, it is possible that the unbound concentrations that are available for SRB1 receptor blockade may have been inadequate to prevent HCV entry. Another aspect of ITX5061 pharmacokinetics that could have contributed to our findings was the plasma concentration profile exhibited after oral dosing. Although a rapid C_{max} was achieved, the decline in plasma concentrations occurred rapidly with a fairly flat concentration profile for the 12-24 hour segment of the dosing period. It is possible that these lower concentrations may have been inadequate to achieve SRB1 receptor blockade throughout the dosing interval and that more frequent dosing (twice or thrice daily) may have been more effective.

Nonetheless, although not directly measured, the expected liver concentration of ITX5061 was expected to be roughly 10-fold higher than plasma, suggesting that the 150 mg daily dose evaluated should have provided more than adequate ITX5061 exposure to assess for anti-HCV activity. As such, insufficient ITX5061 concentration at the 150 mg dose studied is unlikely to completely explain our findings. Despite achieving adequate drug concentration, inadequate duration of exposure represents

another consideration to explain our findings. The rate of turnover of the pool of infected hepatocytes is not known and it is possible that exposures longer than 28 days may be required to meaningfully impact the pool of infected cells.

Another potential explanation for our findings may be that entry inhibition by blocking SR-B1 may be ineffective in the setting of an established, chronic infection. In a humanized mouse model, Ploss and colleagues demonstrated markedly reduced infectivity with a bicistronic HCV genome expressing CRE recombinase in SR-B1 deficient mice, validating the role of this receptor for HCV uptake.(19) However, it is possible that in the context of an established infection, HCV may utilize other hepatocyte receptors to gain entry or that SR-B1 independent cell-to-cell spread of HCV may be an important mechanism for the maintenance of chronic infection and, thus, viral replication may not affected by SR-B1 blockade. Indeed, Catanese and coworkers demonstrated that while SR-B1 was directly involved in cell to cell transmission, the virus was able to lose SR-BI dependence for cell-to-cell spread.(20)

Alternatively, the presence of pre-existing polymorphisms in the HCV E2 glycoprotein which confer resistance to ITX5061 may have led to ineffective blockade of SR-B1. In cell culture-adapted JFH-1 mutants, an amino acid change in E2 at position 415 (N415D) or position 451 (G451R) have been shown to confer resistance to ITX5061 as the result of reduced dependency of the virus on SR-B1 for entry and increased affinity for CD81.(16;21;22) However, these mutations are found in a highly conserved region of E2 downstream of HVR1 suggesting that the presence of high levels of pre-existing variants with reduced SR-B1 dependency is unlikely in our study population (23). Alternatively, it's possible that exposure to ITX5061 monotherapy may have selection for low level pre-existing variants; however, the lack of initial HCV suppression followed by viral rebound argues against the selection of such variants during treatment as a likely explanation for our findings.(16) Interestingly, in a genotype 2a infectious virus system, Zhu and colleague found that ITX5061 was additive to synergistic when given in combination with interferon-alfa, ribavirin, and HCV protease and a nucleoside analogue polymerase inhibitors. As such, in chronically infected patients, ITX5061 in combination with direct acting antivirals may be more effective than ITX5061 monotherapy.

In conclusion, in this proof of concept trial, ITX5061 was safe and well tolerated over 28 days of dosing in non-cirrhotic patients with chronic HCV infection but did not meet predefined criteria for virologic activity. The intriguing finding in one subject of a > 1 log₁₀ decline in HCV viremia coupled with ongoing uncertainty regarding the role, if any, of HCV entry inhibition in chronic infection suggests that additional strategies may warrant further investigation including the combination of putative entry inhibitors with oral HCV direct acting antivirals and their potential use to prevent of new infection following HCV exposure (post-exposure prophylaxis) or recurrent HCV infection following liver transplantation which is currently under investigation (NCT01560468).

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Figure 1. Study design; Safety and virologic results were reviewed after each cohort was complete and prior to the selection of the next cohort. If pre-defined safety criteria were met, the next cohort to enroll was based on the observed virologic activity. Each cohort enrolled 10 new subjects: 8 active, 2 placebo. The cohort sequence enrolled in the study was A150, followed by B150, which was followed by C150 which was final cohort. Following cohort C150, enrollment was terminated according to predefined protocol criteria for observed antiviral activity.

V = pre-defined criteria for virologic activity was observed in the prior cohort. The next cohort enrolled would receive a lower dose of ITX5061.

NV = pre-defined criteria for virologic activity was not observed in the prior cohort. For A150 and B150, the next cohort enrolled would receive the same ITX5061 dose for longer duration. For C150, A75, B75 and C75, the study would end if antiviral activity was not observed.

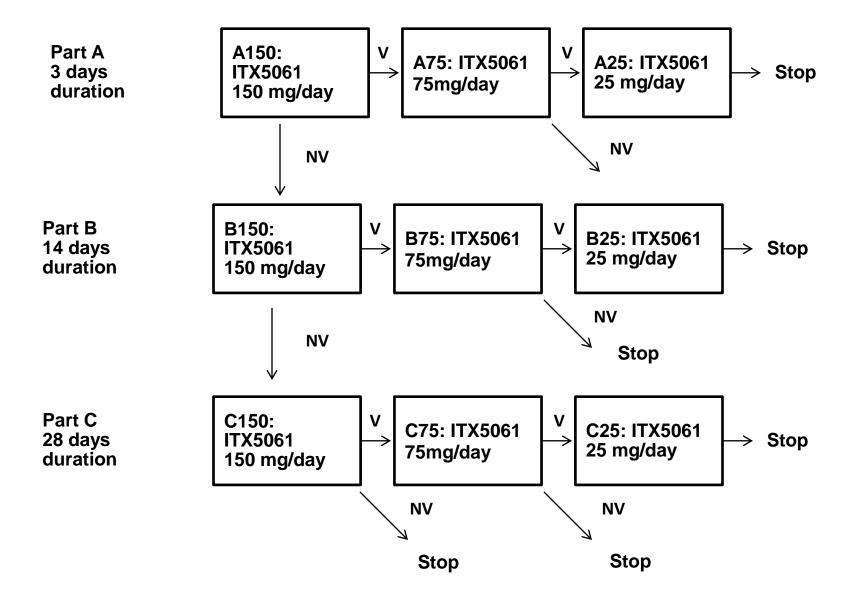
Figure 2. Change in serum log₁₀ HCV RNA level during the dosing interval (● represents the single subject with > 1 log₁₀ decline).

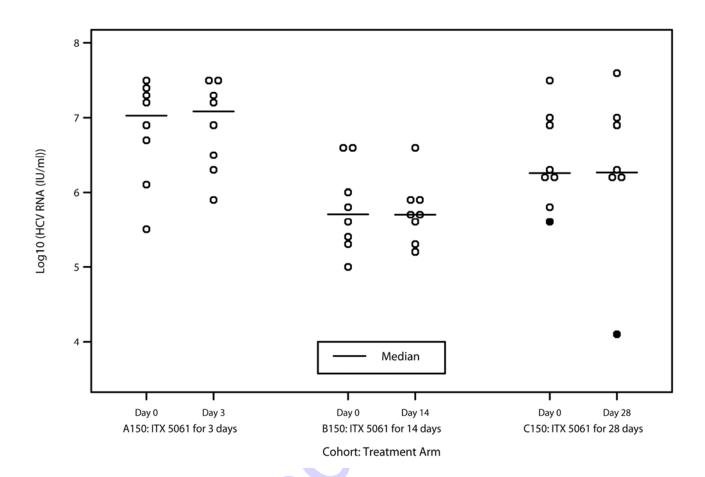
Figure 3. Change in serum log₁₀ HCV RNA and HDL levels over time for the subject dosed with ITX5061 for 28 days (C150) in whom a protocol-defined HCV RNA response was observed.

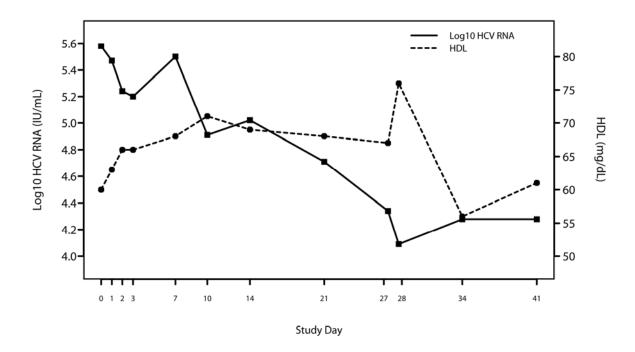
Figure 4. Plasma concentration versus time curves for ITX5061 after 3 (cohort A150), 14 (cohort B150) and 28 (cohort C150) days of dosing at 150 mg by mouth daily (Bars represent interquartile ranges).

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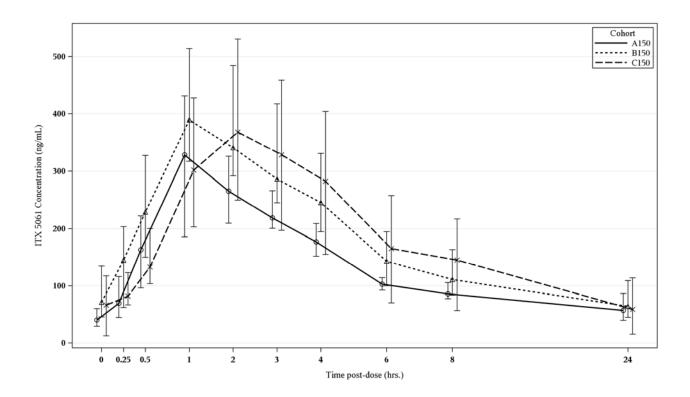




Table 1. Baseline study population characteristics

Characteristic	Cohort A150		Cohort B150)		Cohort C150		Total
	ITX5061	Placebo	ITX5061	Placebo	ITX5061	Placebo	
Number	8	2	8	2	8	2	30
Age, median years (Q1, Q3)	54 (50, 55)	56	46 (37,51)	49	51 (47,54)	47	50 (46, 54)
Female sex	5 (63%)	1 (50%)	5 (63%)	1 (50%)	4 (50%)	0 (0%)	16 (53%)
Race/ethnicity							
White	3 (38%)	1 (50%)	3 (38%)	1 (50%)	1 (13%)	0 (0%)	9 (30%)
Black	4 (50%)	1 (50%)	2 (25%)	1 (50%)	6 (75%)	1 (50%)	15 (50%)
Hispanic	1 (13%)	0 (0%)	3 (38%)	0 (0%)	1 (13%)	1 (50%)	6 (20%)
History of injection drug use	3 (38%)	2 (100%)	6 (75%)	1 (50%)	5 (63%)	1 (50%)	18 (60%)
Weight, median kilograms (Q1, Q3)	88.6 (84.7,98.7)	94.0	94.8 (85.7,104. 1)	79.3	75.3 (65.0, 87.0)	80.6	88.2 (80.2, 100.5)
Log_HCV RNA, median IU/mL, (Q1, Q3)	7.03 (6.39, 7.32)	6.18	5.70 (5.31, 6.28)	5.95	6.26 (5.99, 6.93)	6.41	6.21 (5.72, 6.87)
Serum Alanine Aminotranferase/ULN^, median Q1, Q3)	1.06 (0.62, 2.35)	0.84	1.20 (0.79, 2.02)	1.06	1.04 (0.70, 1.38)	0.80	1.06 (0.65, 1.42)
Direct bilirubin/ULN^, median (Q1, Q3)	0.58 (0.08, 0.97)	0.75	0.50 (0.25 0.83)	0.50	0.50 (0.38, 0.71)	0.75	0.50 (0.33, 0.93)
Indirect bilirubin, median mg/dL (Q1, Q3)	0.30 (0.30, 0.60)	0.45	0.50 (0.25, 0.75)	0.20	0.35 (0.25, 0.65)	0.25	0.30 (0.20, 0.60)
Total cholesterol*, median mg/dL(Q1, Q3)	173 (143, 228)	163	177 (147, 192)	180	176 (152, 205)	147	174 (143. 196)
HDL cholesterol*, median mg/dL(Q1, Q3)	50 (46, 60)	49	53 (47, 65)	51	62 (54, 68)	57	53 (48, 63)
LDL cholesterol*, median mg/dL(Q1, Q3)	88 (80, 131)	100	99 (85, 116)	100	102 (80, 109)	79	99 (79, 115)

[^]ULN= upper limit of normal *Two subjects had non-fasting, baseline lipid profiles.

Table 2. HCV RNA levels prior to (Day 0), at the end of treatment and the change in HCV RNA levels change for subjects dosed with ITX5061 150 mg daily for 3 (A150), 14 (B150) and 28 (C150) days.

Log ₁₀ HCV RNA, IU/mL		Cohort A150	Cohort B150	Cohort C150				
Day 0								
	N	8	8	8				
	Median	7.03	5.70	6.26				
	Q1, Q3	6.39, 7.32	5.31, 6.28	5.99, 6.93				
End of Treatment								
	N	8	8	7				
	Median	7.08	5.70	6.26				
	Q1, Q3	6.36, 7.36	5.49, 5.86	6.20, 6.98				
Change in log ₁₀ HCV RNA from Day 0 at the End of Treatment *								
	N	8	8	7				
	Median	0.06	- 0.04	0.03				
	Q1, Q3	- 0.06, 0.17	- 0.11, 0.08	- 0.14, 0.10				
	P-value [†]	0.46	0.64	0.94				

^{*}p-value= 0.43 by Jonckheere-Terpstra Test for trend across cohorts

[†]Wilcoxon Signed Rank test

Table 3. Plasma concentrations and pharmacokinetic parameters for ITX 5061 after 3 (Cohort A150), 14 (Cohort B150) and 28 (Cohort C150) days of dosing at 150 mg daily*

	Cohort A150 (N=8)	Cohort B150 (N=8)	Cohort C150 (N=7)	p-value [†]
Half Life (hrs)			* . *	0.02
Mean (SD)	29.2 (15.5)	19.1 (5.3)	16.7 (11.4)	
Median (Q1,Q3)	26.7 (18.4, 40.0)	18.5 (17.5, 19.7)	13.4 (8.4, 23.6)	
Tmax (hrs)				0.31
Mean (SD)	1.8 (1.0)	1.5 (0.7)	2.0 (0.5)	
Median (Q1,Q3)	1 (1, 3)	1.1 (1, 2)	2 (2, 2)	
Cmin (ng/mL)				
Mean (SD)	44.3 (19.3)	78.3 (44.5)	66.4 (46.4)	0.22
Median (Q1,Q3)	40 (28.4, 60.3)	61.3 (44.4, 121.7)	66.4 (12.5, 113.3)	
Cmax (ng/mL)				0.28
Mean (SD)	346.6 (116.7)	435.4 (142.4)	419.0 (170.7)	
Median (Q1,Q3)	363.3 (225.6, 431.4)	434.2 (348.4, 515.8)	427.9 (249.7, 531.0)	
C0 (ng/mL)			>	0.22
Mean (SD)	44.5 (19.1)	84.4 (46.6)	68.4 (49.5)	
Median (Q1,Q3)	40.0 (29.1, 60.3)	71.3 (45.6, 134.8)	66.4 (12.5, 117.2)	
C24 (ng/mL)				
Mean (SD)	63.2 (30.2)	74.4 (34.8)	66.6 (44.3)	0.65
Median (Q1,Q3)	57.1 (39.6, 86.6)	64.1 (45.2, 109.2)	59 (15.4, 113.8)	
Clearance (L/hr)				0.18
Mean (SD)	60.2 (16.0)	47.3 (17.3)	53.6 (34.2)	
Median (Q1,Q3)	59.5 (45.7, 70.7)	46.1 (34.8, 56.4)	38.2 (28.2, 90.0)	
AUC (ng*hr/mL)				0.18
Mean (SD)	2644.4 (675.7)	3585.1 (1394.3)	3659.1 (1668.4)	
Median (Q1,Q3)	2527.5 (2133.0, 3294.5)	3268.0 (2665.0, 4486.0)	3929.0 (1667.0, 5320.0)	

Tmax = time at maximum concentration; Cmin = minimum concentration over 24 hours; Cmax = maximum concentration over 24 hours; C0 = pre-dose concentration; C24 = concentration at 24 hours; AUC = area under the concentration-time curve over 24 hours